Serum miRNA profiling identifies miR-150/30a as potential biomarker for workers with damaged nerve fibers from carbon disulfide

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Abstract: As crucial small regulatory molecules, serum microRNAs (miRNAs) have been widely identified as potential noninvasive biomarkers. To survey and identify serum miRNAs associated with workers who had experienced injury to their nerve system from carbon disulfide (CS₂), we profiled abnormally expressed miRNAs using the microarray technique and further performed qRT-PCR validation in case and control samples (n=20). Microarray profiling in pooled RNA samples showed that many miRNAs in workers exposed to CS₂ were aberrantly expressed. Based on control samples exposed to CS₂, a great amount of abnormal miRNAs, including some miRNA gene clusters and families, were obtained from microarray datasets. Most of deregulated miRNAs were up-regulated, and almost all miRNAs showed consistent expression patterns between workers with different numbers of damaged nerve fibers. Functional enrichment analysis suggested that these abnormal miRNAs showed versatile roles by contributing to multiple biological processes. Some aberrantly expressed miRNAs were characterized as miRNA gene clusters or families, and they always showed consistent expression patterns. miR-150 and miR-30a were selected to be further validated by qRT-PCR as up-regulated species, and they could discern case samples from control samples. miR-150 and miR-30a may be potential noninvasive biomarkers for a damaged nervous system.

Key words: microRNA (miRNA), Carbon disulfide (CS₂), Biomarker

Introduction

Carbon disulfide (CS₂) can lead to damage of the nervous system and affect the blood pressure and lipid concentration. A class of small non-coding RNAs (ncRNAs) and microRNAs (miRNAs) have been widely studied as crucial regulatory molecules via targeting mRNAs. miRNAs have important roles in multiple biological processes, including cell development, cell proliferation, apoptosis and differentiation. Simultaneously, some abnormal miRNAs are also involved in pathological processes, even in tumorigenesis.

Although the small ncRNAs have been widely studied,
more studies suggest that some miRNA gene families or clusters may have more versatile biological roles via coordinated regulation patterns. These related miRNAs may have various enrichment levels, but always contribute to multiple biological and pathological processes, including tumorigenesis\(^6\)–\(^8\). A class of special miRNAs in serum tissues, termed circulating miRNAs, may be partly derived from diseased tissues and may be correlated with tumor progression. Novel potential noninvasive blood-based biomarkers have been reported because of the sensitive and informative characteristics\(^9\)–\(^11\).

Here, we attempted to survey and identify miRNAs associated with occupation exposure to CS\(_2\) in workers with damaged nerve fibers. In the study, nerve injury was estimated mainly according to the two ways: demyelination and diffuse axonal injury. The main steps were as follows: [1] serum samples were collected from workers based on the number of damaged nerve fibers (case and control samples), and control serum samples were simultaneously collected from volunteers that were not exposed to CS\(_2\) (independent control samples); [2] miRNA expression profiles in equally pooled serum samples were detected by applying the microarray technique; [3] abnormally expressed miRNAs were screened, and several of them were further identified by qRT-PCR. The study provides data for further selection of noninvasive clinical biomarkers of circulating miRNAs.

### Materials and Methods

Serum samples were collected from workers exposed to CS\(_2\) with 0, 1 and 2–5 damaged nerve fibers, and independent control samples were obtained from volunteers that were not exposed to CS\(_2\) (Table 1). Each group contained 20 patients or volunteers. Other factors, such as age, work age, smoking, drinking, systolic pressure and diastolic pressure were also obtained. Written informed consent was obtained from all the patients and volunteers, and the study was approved by the ethics committee of Nanjing Medical University.

<table>
<thead>
<tr>
<th></th>
<th>Case 1 (20)</th>
<th>Case 2 (20)</th>
<th>Control 1 (20)</th>
<th>Control 2 (20)</th>
<th>(p)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of damaged nerve fibers</td>
<td>2–5</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Age</td>
<td>46.05 ± 4.98</td>
<td>45.45 ± 4.70</td>
<td>44.35 ± 3.79</td>
<td>46.40 ± 5.39</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Work age</td>
<td>21.95 ± 7.72</td>
<td>21.75 ± 6.66</td>
<td>21.25 ± 7.77</td>
<td>23.55 ± 6.83</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Smoking (%)</td>
<td>12 (60%)</td>
<td>14 (70%)</td>
<td>17 (85%)</td>
<td>14 (70%)</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Drinking (%)</td>
<td>2 (10%)</td>
<td>3 (15%)</td>
<td>1 (5%)</td>
<td>4 (20%)</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Systolic pressure (mmhg)</td>
<td>125.95 ± 15.25</td>
<td>128.15 ± 16.73</td>
<td>122.30 ± 15.97</td>
<td>136.00 ± 14.52</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Diastolic pressure (mmhg)</td>
<td>80.10 ± 12.61</td>
<td>84.20 ± 13.75</td>
<td>81.60 ± 10.73</td>
<td>91.10 ± 12.40</td>
<td>&gt;0.05</td>
</tr>
</tbody>
</table>

The ANOVA statistical analysis (\(p\) value) is estimated.
serum samples was isolated with Qiagen miRNeasy Mini kit (Qiagen, Valencia, CA, USA). According to the indicated manufacturer’s instructions, the miRNA bulge-loop was reverse transcribed using the TaqMan miRNA RT Kit and stem-loop RT primers (Applied Biosystems) and quantified by qPCR using TaqMan miRNA probes (Applied Biosystems). The relative enrichment level of miRNA was normalized to snRNA U6. Averages of independent experiments each performed with standard errors were presented.

All the involved statistic analyses were performed using the Statistical Analysis System software (Version 9.1.3, SAS Institute, NC, USA) and R. *P*<0.05 was considered statistically significant, and all tests were two-tailed.

### Results

No significant difference was detected for other factors between case and control samples based on ANOVA analysis (Table 1). Compared to the control 2 (not contact CS2), 168 miRNAs were aberrantly expressed in control 1 (contact CS2) (Fig. S1 and Table S1). Based on miRNA profiles of control 1 and case samples, aberrantly expressed miRNAs were assessed and obtained (Figs. 1 and S1, Table S2). Abundantly and aberrantly expressed miRNAs always showed consistent deregulated expression patterns in case samples. miR-374a was the only miRNA with opposite deregulation patterns between case 1 and case 2 (the log 2 fold change values were 4.36 and −2.79, respectively). Many abnormal miRNAs were prone to be up-regulated (Fig. 1). These deregulated species showed various expression patterns across different samples (Figs 2 and 3). Based on miRNA-mRNA interactions, these abnormal miRNAs had versatile roles in multiple biological pathways including tumorigenesis and signaling pathways (Table 2).

Of these deregulated miRNA species, some were identified as homologous or clustered miRNAs (Figs. 2 and 3). miRNA members in gene clusters and families might show...
various expression levels. Except for the stably expressed members, others always had consistent expression trends (Figs. 2 and 3). For example, the miR-19 gene family and miR-99b gene cluster were both over-expressed.

In order to screen and validate deregulated miRNA species as potential noninvasive biomarkers further, we selected miR-150/miR-30a and performed qRT-PCR validation. Compared to the control group, the two miRNAs were identified as up-regulated miRNAs (4–5 folds), and they had consistent expression patterns in the two diseased groups. Both of them were identified as associated miRNAs with central nerves system. The qRT-PCR results showed similar expression patterns as observed in bioinformatics analysis (Figs. 4A). The ROC curve indicated that the two miRNAs could discern damaged nerve fibers cases from control samples (Fig. S2). These validated up-regulated miRNAs have important roles in multiple biological processes, including O-Glycan biosynthesis and Axon guidance, etc. (Fig. 4B).

Discussion

Microarray data showed a series of aberrantly expressed miRNAs, including some miRNA gene clusters and families (Tables S1 and S2, Figs. 1–3). Compared to workers that were not exposed to CS₂, many miRNAs were aberrantly expressed in control 1 (Table S1). These results indicate that CS₂ may directly or indirectly regulate expression levels of miRNAs. The detailed mechanisms should be derived from the toxic mechanism of CS₂. For the exposed workers, almost all miRNAs have consistent up- or down-regulated expression patterns with different numbers of damaged nerve fibers (Figs. 1 and S1). Interestingly, in these differentially expressed miRNAs, the up-

Fig. 2. The dynamic expression patterns of homologous miRNAs across different pooled serum samples based on ΔC_T (the vertical coordinates). All the homologous members in the specific miRNA gene family are presented here, although some are stably expressed. Some homologous miRNAs are also clustered in specific genomic region. The ΔC_T reflects the relative expression levels of miRNAs, i.e., the higher the ΔC_T value the lower the expression level.
Fig. 3. The dynamic expression patterns of clustered miRNAs across different pooled serum samples based on ΔC_T (the vertical coordinates). All of these involved miRNAs are aberrantly expressed in case samples.

Fig. 4. qRT-PCR validation and functional enrichment analysis. (A) qRT-PCR validation; (B) functional enrichment analysis.
regulation patterns are more popular, which indicates that potential target mRNAs are negatively regulated by these up-regulated miRNAs. Aberrant expression of mRNAs may lead to abnormal biological pathways and even pathological processes. Based on validated target mRNAs of deregulated miRNAs, the functional enrichment analysis showed that these miRNAs have important roles in multiple biological processes, including some cancers (Table 2).

Observed abnormal miRNAs may have close physical or sequence relationships, and they always are characterized as miRNA gene clusters and families (Figs. 2 and 3). These miRNAs may co-regulate multiple biological processes22–24, such as the versatile roles of the miR-17-92 gene cluster and family6–8). Clustered or homologous miRNAs may have various enrichment levels in case samples, although some clustered miRNAs may be co-transcribed22, 23, 25. Some members may be stably expressed, while others always show consistent aberrant expression patterns (Figs. 2 and 3). The consistent expression trends and the expression patterns may contribute to coordinated regulation between different miRNAs. The synergistic interaction further enriches the miRNA regulatory networks and miRNA-mRNA interaction. Based on the potential interaction between different miRNAs, miRNA gene clusters and families may be novel biomarkers for diagnosing diseases. The dynamic expression patterns with consistent deregulation patterns suggest their important roles in abnormal biological processes and imply their potential characters as candidate biomarkers.

Based on differentially expressed miRNAs from microarray datasets, qRT-PCR further validated up-regulated miR-150 and miR-30a in case 1, case 2 and control 1 samples (n=20, Figs. 4A and S2). miR-30a may function as a metastasis suppressor in metastatic colorectal carcinoma26 and may be a tumor-suppressing miRNA in colon cancer cells27. Indeed, another miRNA, miR-150 also has versatile biological roles. Serum miR-150 may have an important role in the pathogenesis of systemic sclerosis via regulating integrin beta328. Based on experimentally validated target mRNAs, functional enrichment analysis shows that the two miRNAs have versatile roles in multiple biological pathways (Fig. 4B). The results show that CS2 can directly or indirectly lead to aberrant expression of miRNAs through the complex toxic mechanisms. These deregulated small RNAs further contribute to damage of the nervous system. As flexible small regulatory molecules, miRNAs have important roles in many biological processes, including cell development, cell proliferation, apoptosis and differentiation4, 5). Therefore, miRNAs may be important mediums between CS2 and the damaged nervous systems and could be potential noninvasive biomarkers.

Acknowledgements

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References


Appendix

Table S1. The top 10 up- and down-regulated miRNA species in control 1

<table>
<thead>
<tr>
<th>Up-regulated miRNAs</th>
<th>FC (abs)</th>
<th>Down-regulated miRNAs</th>
<th>FC (abs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>miR-151-3p</td>
<td>1,621.37</td>
<td>miR-409-3p</td>
<td>1026.13</td>
</tr>
<tr>
<td>miR-539</td>
<td>140.75</td>
<td>miR-28-3p</td>
<td>676.99</td>
</tr>
<tr>
<td>miR-518b</td>
<td>91.27</td>
<td>miR-324-3p</td>
<td>582.45</td>
</tr>
<tr>
<td>miR-433</td>
<td>32.49</td>
<td>miR-223*</td>
<td>535.60</td>
</tr>
<tr>
<td>miR-198</td>
<td>26.46</td>
<td>miR-146b-5p</td>
<td>478.04</td>
</tr>
<tr>
<td>miR-101</td>
<td>25.79</td>
<td>miR-425</td>
<td>465.62</td>
</tr>
<tr>
<td>miR-520h</td>
<td>20.15</td>
<td>miR-155</td>
<td>386.68</td>
</tr>
<tr>
<td>miR-181a-2*</td>
<td>19.20</td>
<td>miR-20b</td>
<td>356.07</td>
</tr>
<tr>
<td>miR-200a*</td>
<td>13.75</td>
<td>miR-106b</td>
<td>342.75</td>
</tr>
<tr>
<td>miR-769-5p</td>
<td>12.54</td>
<td>miR-10b*</td>
<td>333.61</td>
</tr>
</tbody>
</table>

These deregulated miRNA species are assessed using absolute fold change (abs) based on control 2.

Table S2. The top 10 up- and down-regulated miRNA species

<table>
<thead>
<tr>
<th>Up-regulated miRNAs</th>
<th>FC (abs)</th>
<th>Down-regulated miRNAs</th>
<th>FC (abs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>miR-425</td>
<td>1,105.13</td>
<td>miR-151-3p</td>
<td>829.44, 871.28</td>
</tr>
<tr>
<td>miR-324-3p</td>
<td>805.64, 127.56</td>
<td>miR-539</td>
<td>181.90, 253.88</td>
</tr>
<tr>
<td>miR-28-3p</td>
<td>800.63, 253.53</td>
<td>miR-518b</td>
<td>169.13, 164.62</td>
</tr>
<tr>
<td>miR-146b-5p</td>
<td>436.85, 72.86</td>
<td>miR-152</td>
<td>80.39, 0.91</td>
</tr>
<tr>
<td>miR-155</td>
<td>376.11, 28.78</td>
<td>miR-513-3p</td>
<td>56.61, 75.58</td>
</tr>
<tr>
<td>miR-20b</td>
<td>363.04, 117.87</td>
<td>miR-590-5p</td>
<td>36.45, 0.83</td>
</tr>
<tr>
<td>miR-132</td>
<td>333.14, 47.87</td>
<td>miR-596</td>
<td>35.33, 47.18</td>
</tr>
<tr>
<td>miR-223*</td>
<td>300.25, 67.00</td>
<td>miR-708</td>
<td>30.93, 30.11</td>
</tr>
<tr>
<td>miR-145</td>
<td>297.14, 19.31</td>
<td>miR-101</td>
<td>28.40, 46.53</td>
</tr>
<tr>
<td>miR-409-3p</td>
<td>274.18, 221.48</td>
<td>miR-148b</td>
<td>27.83, 27.10</td>
</tr>
</tbody>
</table>

These deregulated miRNA species are assessed based on absolute fold change (abs). The top 10 miRNAs were collected based on case 1 with 2–5 damaged nerve fibers. The abs values of case 2 were also presented.
Fig. S1. The number distributions of deregulated miRNA species in case 1 and case 2 (based on control 1), and control 1 (based on control 2). (A) Number of significantly up-regulated and down-regulated miRNAs between case 1, case 2 and control 1; (B) Number of down-regulated miRNA species between case 1 and case 2; (C) Number of up-regulated miRNA species between case 1 and case 2.
Fig. S2. ROC curve analysis for discrimination between case 1, case 2 and control 1 samples by the miR-150 and miR-30a.